

JURNAL TEKNOLOGI LABORATORIUM

Home / Archives / Vol 9 No 1 (2020): 2020 (1): Special Edition "COVID-19"



In this section, we published a special issue about COVID-19. All papers in this number have been reviewed with all our reviewers. We hope these papers will be useful for cope with COVID-19.

DOI: https://doi.org/10.29238/teknolabjournal.v9i1

 Modulation of severe acute respiratory syndrome coronavirus (SARS-CoV-2)
 Intraceives martin

 Indra Lasmana Tarigan, Kartika Arum
 Only received

 Indra Lasmana Tarigan, Kartika Arum
 English La

 Image: Dol 10.29238/teknolabjournal.v9i1.214
 Submission Proceived

 Immunobioinformatics analysis and phylogenetic tree construction of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) in Indonesia: spike glycoprotein gene
 Editorial Team

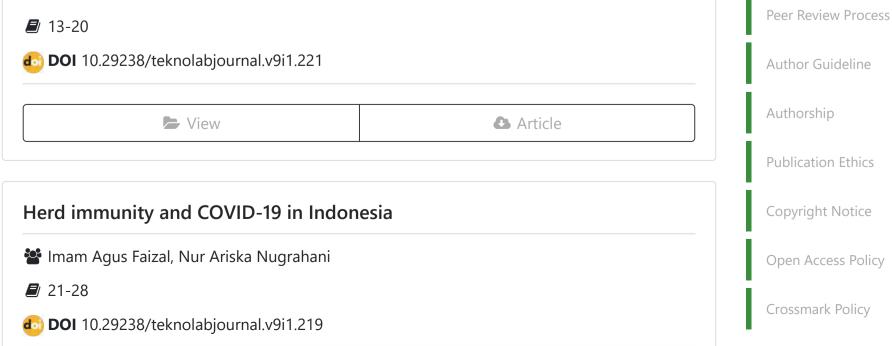
Arif Nur Muhammad Ansori, Viol Dhea Kharisma, Yulanda Antonius, Martia Rani Tacharina, Fedik Abdul Rantam

J-Jalk



Call for Papers JTL receives manuscripts at any time
Only received Manuscript in English Language
Submission Procedure
Focus and Scope
Editorial Team
List of Reviewers

Reviewer Guideline



View		Article	Plagiarism Screening
			Publication Fee
Human immune respon	ise to SARS-Co	V-2 infection	Publication Frequency
📽 Lia Yosaneri Wina Nurtias	, Dora Dayu Rahma	a Turista, Eka Puspitasari	Archiving
2 9-40			Journal Indexing
둴 DOI 10.29238/teknolabjo	urnal.v9i1.223		
			Author Index
View		🕰 Article	Ethical Approval
 Ozüdogru Osman, Gunes 41-48 DOI 10.29238/teknolabjo 			Citedness in
	umai.v911.220		
View		🕰 Article	Scopu
Indonesia		avirus (COVID-19) outbreak in	CITATION ANALYSIS
Sutaryono Sutaryono, Sho 49-57	olikhah Deti Andasa	ari, Heru Subaris Kasjono	GS Citation: JTL : GS Citation: JTL
DOI 10.29238/teknolabjo	urnal.v9i1.222		All
<u> </u>			Citations h-index
View		🕰 Article	i10-index
> View		Article	GS Citation: JTL
	ır immune resp	Article onse of SARS-COV-2 infection	
	•	onse of SARS-COV-2 infection	
Perspective of molecula	•	onse of SARS-COV-2 infection	GS Citation: JTL
Perspective of molecula	Arif Rahman Nurdia	onse of SARS-COV-2 infection	

🕨 View

Article

Indirect exposure to novel coronavirus (SARS-CoV-2): an overview of current knowledge

Christian Ebere Enyoh, Andrew Wirnkor Verla, Wang Qingyue, Dipak Kumar Yadav, Md Akhter Hossain Chowdhury, Beniah Obinna Isiuku, Tanzin Chowdhury, Francis Chizoruo Ibe, Evelyn Ngozi Verla, Tochukwu Oluwatosin Maduka

┛ 67 - 77

do	DOI	10.29238/teknolabjournal.v9i1.227	
----	-----	-----------------------------------	--

> View	🕰 Article	



MOST READ LAST WEEK

Tweets by @JurnalTeknolog1

Jurnal Teknologi Laboratorium

Comparison of real-time reverse transcriptase polymerase chain reaction	
(RT-PCR) and IgM and IgG antibody test for the diagnosis of SARS-CoV-2	
infection	

Ömer ACER, Osman ÖZÜDOĞRU

2 78-86

DOI 10.29238/teknolabjournal.v9i1.232

View

Article



DOI 10.29238/teknolabjournal.v9i1.230

b View

Article





Some #journals say that they are indexed in DOAJ but they are not.docs.google.com/spr eadsheets/d...

Never take what you read on a journal homepage at face value. Use **#DOAJ** to check and use tools like @thinkchecksub #ThinkCheckSubmit #AcademicTwitter #schlcomm #Openaccess



Jan 27, 2021



Jurnal Teknol @JurnalTekno

The most important thing in our journal is a substantial factor... check it out at teknolabjournal.com



Jan 23, 2021



Jurnal Teknol @JurnalTekno

All research need ethical approval? See our policy at teknolabjournal.com/inde x.php/Jtl/...





Jan 12, 2021

Jurnal Teknologi Laboratorium Retweeted

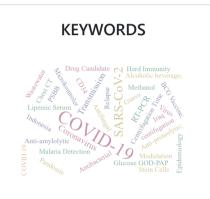


.@SussexUni has embarked upon an institutional project endorsed by the Heads of all academic Schools, to build on existing good practice in research assessment & to ensure that these principles are implemented and

embedded across the institution.sussex.ac.uk/r esearch/about...

<section-header><section-header><section-header><section-header><section-header><section-header><section-header><section-header>









JURNAL TEKNOLOGI LABORATORIUM

Jurusan Analis Kesehatan

Poltekkes Kemenkes Yogyakarta

Jl. Ngadinegaran MJ III/62 Yogyakarta, Indonesia

ISSN 2338-5634 (Print)

ISSN 2580-0191 (Online)

email: budi.setiawan@poltekkesjogja.ac.id

Location:



INFORMATION
• For Readers
• For Authors
• For Librarians







JURNAL TEKNOLOGI LABORATORIUM by Poltekkes Kemenkes Yogyakarta

is licensed under a Creative Commons Attribution-ShareAlike 4.0 International License.

About Jurnal Teknologi Laboratorium

Disclaimer: Articles published by Jurnal Teknologi Laboratorium have been pre-viewed and authenticated by the Authors before publication. The Journal, Editor and the editorial board are

Website Links	Journal Links	Follow Us
About	» Issue	f Facebook
🗹 Contact	» Archive	Google+

1/29/2021

Vol 9 No 1 (2020): 2020 (1): Special Edition "COVID-19" | Jurnal Teknologi Laboratorium

responsible for inaccurate and misleading data if any. Read our Plagiarism Policy and use of this site signifies your agreement to the Terms of Use

 Submissions 	>>>	Current	y	Twitter
🐮 Editor	\gg	Search	O)	Instagram
Team				

Copyright \bigcirc 2018 Jurnal Teknologi Laboratorium – All rights reserved System by Open Journal Systems 3.0

Themes by Informatics Engineering UIN SGD Bandung Designed with ♥ and 里

JURNAL TEKNOLOGI LABORATORIUM

A Home / Editorial Team

Editorial Team

EDITOR-IN-CHIEF

Budi Setiawan (ORCID ID) (Scopus ID 57215207015) (Department of Medical Laboratory Technology, Poltekkes Kemenkes Yogyakarta, Indonesia)

Regional Editors for Asia Region

Sirisha Kanugala (Scopus ID 57207915232) (Department of Biotechnology Senior research fellow- CSIR-Indian Institute of Chemical Technology, India)

Dipak Kumar Yadav (Orcid ID 0000-0003-4427-1964) (Nobel Medical College Teaching Hospital, Biratnagar, Nepal)

Regional Editors for Africa Region

Christian Ebere Enyoh (Scopus ID 57209317908) (Department of Chemistry, Imo State University (IMSU), Imo State, Nigeria)

Regional Editors for Europe Region

Gunes Bolatli (Orcid ID 0000-0002-7648-0237) (Department of Anatomy, Faculty of Medicine, Siirt University, Turkey)

Associate Editors for Other Regions

Anis Nurwidayati (Balai Litbang P2B2 Donggala Kemenkes RI, Indonesia)

Anna Yuliana (Scopus ID: 57192715936) (STIKES Bakti Tunas Husada, Tasikmalaya, Indonesia)

Dora Dayu Rahma Turista (Scopus ID: 57217389770) (STIKes Hutama Abdi Husada Tulungagung, Indonesia)





Call for Papers
JTL receives manuscripts at any time
Only received Manuscript in English Language
Submission Procedure
Focus and Scope
Editorial Team
List of Reviewers
Reviewer Guideline

Peer Review Process
Author Guideline
Authorship
Publication Ethics
Copyright Notice
Open Access Policy
Crossmark Policy

Plagiarism Screening

Publication Fee

Publication Frequency

Archiving

Journal Indexing

Author Index

Ethical Approval



Citedness in Scopus*

CITATION ANALYSIS

GS Citation: JTL : GS Citation: JTL

> Let's change what we value in research.



MOST READ LAST WEEK

Tweets by @JurnalTeknolog1

Jurnal Teknologi Laboratorium

Retweeted



@DOAJplus

Some #journals say that they are indexed in DOAJ but they are not.docs.google.com/spr eadsheets/d...

Never take what you read on a journal homepage at face value. Use **#DOAJ** to check and use tools like @thinkchecksub #ThinkCheckSubmit #AcademicTwitter #schlcomm #Openaccess



Jan 27, 2021



Jurnal Teknol @JurnalTekno

The most important thing in our journal is a substantial factor... check it out at teknolabjournal.com



Jan 23, 2021



Jurnal Teknol @JurnalTekno

All research need ethical approval? See our policy at teknolabjournal.com/inde x.php/Jtl/...

•		#	1 4 9
Type of study	Study design	informed content	Ethical approval
Research.	The moterial from patients/healthy forors in collected for research perpose		Required
	This use of residual material	Not required	Regared
Method/instrument validation	The material from patients/healthy denors in collected for research	Required	Regared



Jan 12, 2021

Jurnal Teknologi Laboratorium Retweeted



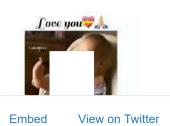
.@SussexUni has embarked upon an institutional project endorsed by the Heads of all academic Schools, to build on existing good practice in research assessment & to ensure that these principles are implemented and

embedded across the institution.sussex.ac.uk/r esearch/about... Jan 11, 2021

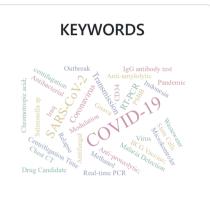
Jurnal Teknologi Laboratorium Retweeted



Love you.











JURNAL TEKNOLOGI LABORATORIUM

Jurusan Analis Kesehatan

Poltekkes Kemenkes Yogyakarta

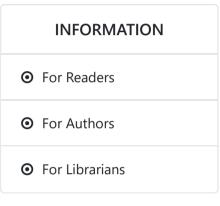
Jl. Ngadinegaran MJ III/62 Yogyakarta, Indonesia

ISSN 2338-5634 (Print)

ISSN 2580-0191 (Online)

email: budi.setiawan@poltekkesjogja.ac.id

Location:









JURNAL TEKNOLOGI LABORATORIUM by Poltekkes Kemenkes Yogyakarta

is licensed under a Creative Commons Attribution-ShareAlike 4.0 International License.

About Jurnal Teknologi Laboratorium

Disclaimer: Articles published by Jurnal Teknologi Laboratorium have been pre-viewed and authenticated by the Authors before publication. The Journal, Editor and the editorial board are

Website Links	Journal Links	Follow Us
About	» Issue	f Facebook
🗹 Contact	» Archive	Google+

1/29/2021

Editorial Team | Jurnal Teknologi Laboratorium

responsible for inaccurate and misleading data if any. Read our Plagiarism Policy and use of this site signifies your agreement to the Terms of Use

 Submissions 	» Current	Y Twitter
😫 Editor	» Search	Instagram
Team		

Copyright © 2018 Jurnal Teknologi Laboratorium – All rights reserved System by Open Journal Systems 3.0 Themes by Informatics Engineering UIN SGD Bandung Designed with 🎔 and 🖭

https://www.teknolabjournal.com/index.php/Jtl/about/editorialTeam

JURNAL TEKNOLOGI LABORATORIUM

List of Reviewer

Agus Wijanarka (Scopus ID 57190808066) (Department of Nutrition, Poltekkes Kemenkes Yogyakarta)

Arif Nur Muhammad Ansori (Scopus ID 57195995342) (Airlangga University, Surabaya, Indonesia)

Arli Aditya Parikesit (Scopus ID 26531908500) (Bioinformatics Departement, Indonesia International Institute for Life Science)

Jusak Nugraha (Scopus ID 46662009200) (Airlangga University, SUrabaya, Indonesia)

Kuntaman (Scopus Id: 8700386400) (Faculty of Medicine, Universitas Airlangga, Surabaya, Indonesia)

Mahde Saleh Assafi (Scopus ID 55331393500) (University of Duhok, Iraq)

Mohammad Rizki Fadhil Pratama (Scopus id: 56925239400) (Faculty of Pharmacy, Universitas Muhammadiyah Palangkaraya, Indonesia)

Muji Rahayu (Department of Medical Laboratory Technology, Poltekkes Kemenkes Yogyakarta, Indonesia)

Nanik Sulistyani (Scopus ID 57096330300) (Faculty of Pharmacy, Universitas Ahmad Dahlan, Yogyakarta, Indonesia)

Ni Kadek Warditiani (Scopus ID 55241768500) (Faculty of Pharmacy, Udayana University, Bali, Indonesia)

Rahayu Anggraini (Department of Medical Laboratory Technology, Universitas Nahdlatul Ulama Surabaya, Indonesia)

Serena Cavallero (Department of Parasitology, University of Sapienza, Rome, Italy)

Sirisha Kanugala (Scopus ID 57207915232) (Department of Biotechnology Senior research fellow-CSIR-Indian Institute of Chemical Technology)

Ummy Mardiana Ramdan (Scopus ID 57039105800) (STIKes Bakti Tunas Husada Tasikmalaya)

Made Ary Sarasmita (Scopus ID 57200126381) (Department of Clinical Pharmacy, School of Pharmacy, Taipei Medical University, Taipei, Taiwan)

J-Jalk



Call for Papers
JTL receives manuscripts at any time
Only received Manuscript in English Language
Culturationian Decondum
Submission Procedure
Focus and Scope
Editorial Team
List of Reviewers
Reviewer Guideline

Peer Review Process

Author Guideline

Publication Ethics

Copyright Notice

Open Access Policy

Crossmark Policy

Authorship

https://www.teknolabjournal.com/index.php/Jtl/rev

Plagiarism Screening

Publication Fee

Publication Frequency

Archiving

Journal Indexing

Author Index

Ethical Approval



Citedness in Scopus*

CITATION ANALYSIS

T.

GS Citation: J Citation: JTL	TL : GS
	All
Citations	193
h-index	8
i10-index	6
GS Citation: JTL	>

Let's change what we value in research.



MOST READ LAST WEEK

Tweets by @JurnalTeknolog1

Jurnal Teknologi Laboratorium

Retweeted



@DOAJplus

Some #journals say that they are indexed in DOAJ but they are not.docs.google.com/spr eadsheets/d...

Never take what you read on a journal homepage at face value. Use **#DOAJ** to check and use tools like @thinkchecksub #ThinkCheckSubmit #AcademicTwitter #schlcomm #Openaccess



Jan 27, 2021



@JurnalTekno

The most important thing in our journal is a substantial factor... check it out at teknolabjournal.com



Jan 23, 2021



Jurnal Teknol @JurnalTekno

All research need ethical approval? See our policy at teknolabjournal.com/inde x.php/Jtl/...

•		II Q 4 0		
Type of study	Study design	informed	Ethical approval	
Research.	The moterial from patients/hea/thy donors in collected for research perpose	Required	Required	
	The use of residual material	Not required	Regared	
Nethod/instrument validation	The material from patients/healthy donors is collected for research	Required	Regared	



Jan 12, 2021

Jurnal Teknologi Laboratorium Retweeted

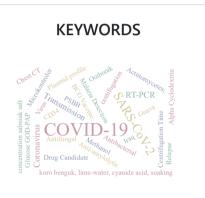


.@SussexUni has embarked upon an institutional project endorsed by the Heads of all academic Schools, to build on existing good practice in research assessment & to ensure that these principles are implemented and

embedded across the institution.sussex.ac.uk/r esearch/about...











JURNAL TEKNOLOGI LABORATORIUM

Jurusan Analis Kesehatan

Poltekkes Kemenkes Yogyakarta

Jl. Ngadinegaran MJ III/62 Yogyakarta, Indonesia

ISSN 2338-5634 (Print)

ISSN 2580-0191 (Online)

email: budi.setiawan@poltekkesjogja.ac.id

Location:









JURNAL TEKNOLOGI LABORATORIUM by Poltekkes Kemenkes Yogyakarta

is licensed under a Creative Commons Attribution-ShareAlike 4.0 International License.

About Jurnal Teknologi Laboratorium

Disclaimer: Articles published by Jurnal Teknologi Laboratorium have been pre-viewed and authenticated by the Authors before publication. The Journal, Editor and the editorial board are

Website Links	Journal Links	Follow Us		
About	» Issue	f Facebook		
🗹 Contact	» Archive	Google+		

1/29/2021

List of Reviewer | Jurnal Teknologi Laboratorium

responsible for inaccurate and misleading data if any. Read our Plagiarism Policy and use of this site signifies your agreement to the Terms of Use

 Submissions 	» Current	Y Twitter
Editor	» Search	Instagram
Team		

Copyright @ 2018 Jurnal Teknologi Laboratorium – All rights reserved System by Open Journal Systems 3.0

Themes by Informatics Engineering UIN SGD Bandung Designed with 🎔 and 🖭

SERTIFIKAT

Direktorat Jenderal Penguatan Riset dan Pengembangan, Kementerian Riset, Teknologi dan Pendidikan Tinggi





Kutipan dari Keputusan Direktur Jenderal Penguatan Riset dan Pengembangan Kementerian Riset, Teknologi dan Pendidikan Tinggi Republik Indonesia Nomor: 23/E/KPT/2019 PERINGKAT AKREDITASI JURNAL ILMIAH PERIODE IV TAHUN 2019 Nama Jurnal Ilmiah

Jurnal Teknologi Laboratorium

E-ISSN: 25800191

Penerbit: Poltekkes Kemenkes Yogyakarta

Ditetapkan Sebagai Jurnal Ilmiah

TERAKREDITASI PERINGKAT 2

Akreditasi Berlaku Selama 5 (lima) Tahun, Yaitu





Article Review



Perspective of molecular immune response of SARS-COV-2 infection

Martina Kurnia Rohmah^{1*}, Arif Rahman Nurdianto²

¹ Department of Pharmacy, STIKES Rumah Sakit Anwar Medika, Sidoarjo, Indonesia ² Public Health Office of Sidoarjo, Sidoarjo, Indonesia

¹E-mail address : <u>martina.kurniarohmah@gmail.com</u> ²E-mail address : <u>didins99@gmail.com</u>

HIGHLIGHTS

SARS-CoV-2 increases the number of neutrophils as APC, suppresses IFN, increases the activity of Th1 / Th17, B cells, CD8 + and CD4 +, and causes cytokine storms especially pro-inflammatory cytokines which can increase respiration disorders and multi-organ damage.

ARTICLE INFO

Article history:

Received Date: May 07, 2020 Revised Date: June 04, 2020 Accepted Date: June 18, 2020

Keywords:

COVID-19 SARS-CoV-2 Immune Response

ABSTRACT

COVID-19 is a type of Pneumonia that caused by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). When COVID-19 arise in Wuhan China and rapidly spread throughout to the World, we need to learn how pathogenesis and immune responses occur in the bodies in more detail. COVID-19 is third Severe Respiratory Disease outbreak caused by the Coronavirus in the past two decades after Severe Acute Respiratory Syndrome (SARS) in the 2002 and Middle East Respiratory Syndrome (MERS) in the 2012. The Articles from PUBMED and Research Gate were searched for studies on the immune response of COVID-19 infection by SARS-CoV-2. SARS-CoV-2 increases the number of neutrophils, suppresses IFN, increases the activity of Th1/Th17, B cells, CD8+ and CD4+, and causes cytokine storms especially pro-inflammatory cytokines which can increase respiration disorders and multi-organ damage. This review tries to explain about pathogenesis and immune responses of COVID-19 to provide a reference in designing the appropriate immune intervention for treatment and therapeutic such as drug or vaccine based on the recent research progress SARS-CoV-2 and previous studies about SARS CoV and MERS CoV.

This is an open-access article under the CC-BY-SA license.

 \odot \odot \odot

*Corresponding Author: Martina Kurnia Rohmah, Department of Pharmacy, STIKES RS Anwar Medika, Sidoarjo, Indonesia JI. Raya By Pass Krian KM 33, Sidoarjo, Jawa Timur, Indonesia Email: martina.kurniarohmah@gmail.com Phone: +6285859686848



1. INTRODUCTION

Coronavirus Disease 2019 (COVID-19) is a pathogenic viral infection caused by Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) that initially arises in Wuhan, Hubei Province, China and rapidly spread to the world. Initially, the cases arise in Wuhan's Hunan Seafood Wholesale Market which trade in fish and variety of live animals such as bats, marmots, and snake. The causative agent for the disease was identified from throat swab samples conducted by the Chines Centre for Disease Control and Prevention (CCDC), and was subsequently named Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2). The sequences-based analysis suggested that bat is a key reservoir of SARS-CoV-2 and can be transmitted to humans.¹ Furthermore, the transmission will occur from human to human as it is today.

On 30th January 2020, the World Health Organization (WHO) declared that COVID-19 from Wuhan was to be a Public Health Emergency of the International Concern posing a high risk to countries. WHO announces this disease outbreak a pandemic diseases. It is due to the rapid increase in the number of cases outside China over the past 2 weeks that has affected a growing number of countries in the world.² There were 7734 cases have been confirmed in China on that day. In a number of counties also confirmed 90 cases.³ On 30th March 2020, WHO confirmed that there were 638.146 cases with 30105 death in 203 countries in the World.⁴ In Indonesia, Ministry of Health Indonesia confirmed that there are 1285 positive cases with 114 deaths and 64 recovering on 30th March 2020.⁵

COVID-19 symptoms appear after an incubation period of approximately 5.2 days. The period from the onset of the symptoms to death ranges is from 6 to 41 days with a median of 14 days.^{6.7} This period is dependent on the age of the patient and the status of the immune system. The most clinical symptoms at the onset of COVID-19 are fever, dry cough, and fatigue. The other symptoms include headache, sputum production, hemoptysis, diarrhea, dyspnea, lymphopenia, and bilateral lung infiltrates on imaging.⁷

The main problems of COVID-19 are high transmission and the absence of specific drugs and vaccines. For this problem, the immune system is an important aspect to support the host's self-defense and the patient's recovery. This review may help in designing the appropriate immune intervention for treatment and therapeutic such as drug or vaccine in preventing more wide transmission and in the development of the COVID-19 drugs based on the recent research progress SARS-CoV-2 and previous studies about SARS -CoV, and MERS-CoV.

2. REVIEW METHOD

We performed this review of the Pubmed and Research Gate databases from inception to 30 March 2020 to find articles providing information on the molecular immunopathogenesis of COVID-19 infection. The initial search identified 87 sources. Based on the inclusion criteria, we get 13 articles that discuss the Immunopathogenesis of Coronavirus including SARS-CoV, MERS-CoV, and SARS-CoV-2. Initially, this review explains the immune response in SARS-CoV and MERS-CoV, and subsequently, explains the pathogenesis and immune response in SARS-CoV-2. As a coronavirus family, we assume that the immune response of SARS-CoV-2 is similar to SARS-CoV and MERS-CoV. The search was expanded using a snowballing method applied to the references of retrieved papers.

3. RESULTS AND DISCUSSION

As the coronavirus family, the immune response of SARS-CoV-2 is similar to SARS-CoV and MERS-CoV, although it has little difference in clinical manifestation. The summary of the immune response is described in <u>table 1</u>.

Table 1. I	mm	une Response Summary of SARS-CoV, MERS-CoV, and SARS-C	oV-2
Coronaviruses		Immune Responses	Source
SARS-CoV	1.	Cytotoxic T Lymphocyte (CTL)	[<u>8</u>]
	2.	MHC class I and II (HLA)	
	3.	HLA-B * 4601, HLA-B * 0703, HLA-DR B1 * 1202	[<u>8]</u> [<u>9</u>]
	4.	HLA-Cw * 0801	[<u>10</u>]
	5.	HLA-DR0301, HLA-Cw 1502 and HLA-A * 0201	[<u>11</u>]
	6.	CD8 + T cells	[<u>12</u>]
	7.	Cytokine (IL-1 β , IL-6, IL-12, IL-18, IL-33, TNF- α , TGF- β) and Chemokines (CCL2, CCL3, CCL5, CXCL8, CXCL9, CXCL10)	[<u>38</u> , <u>39</u> , <u>40</u>]

Coronaviruses		Immune Responses	Source
MERS-CoV	1.	MHC class II molecules: HLA-DRB1 * 11: 01 and HLA-DQB1 * 02: 0	[<u>13</u>]
	2.	Mannose Binding Lectine (MBL) gene polymorphism associated with APC	[14]
	3.	CD8+ T cells	[<u>15</u>]
SARS-CoV-2	1.	Neutrofil as APC	[<u>15</u>]
	2.	High Proinflammatory cytokine: IL6, IP-10, MCP-1, MIP-1A, and TNFα.2	[<u>15</u>]
	3.	Th1/Th17	[15]
	4.	Low Antiinflammatory cytokine: Type I IFN	[15]
	5.	B cell, IgM, and IgG	[16]
	6.	CD4+ CD8+ T cells	[17]

Immune response to SARS-CoV-2 infection based on clinical manifestation approach, clinical laboratory test results, and studies of immune response on SARS-CoV and MERS-CoV can be shown in <u>figure 1</u>.

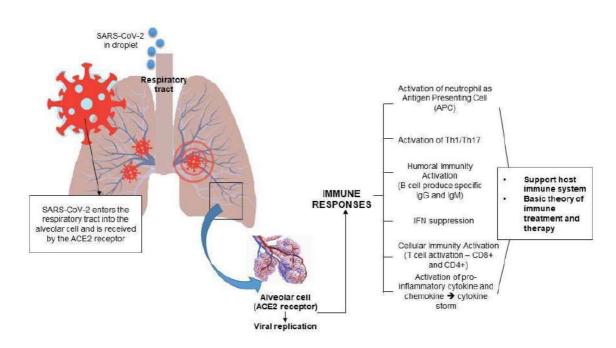


Figure 1. Host immune responses prediction during SARS-CoV-2 infection

According to genotypically and serologically, coronavirus is divided into 4 genera such as α , β , γ , and δ Coronaviruses. The subfamily α and β CoV usually causes Human CoV infections. SARS CoV and MERS CoV is included in β CoV. The SARS-CoV-2 is novel β CoV was named by the International Virus Classification Commission.^{18,16}

Coronaviruses (CoV) are enveloped viruses, non-segmented, and positive-sense singlestranded RNA (ssRNA) virus genomes in the size 26 to 32 kb, the largest known viral RNA genome. SARS-CoV-2 had a length of 29.9 kb. SARS-CoV-2 has four main structural proteins including Spike Glycoprotein (S), small envelope glycoprotein (E), membrane glycoprotein (M), and nucleocapsid protein (N), and several accessory protein. The structure of CoV virion is shown in <u>figure 1</u>.¹⁶

The Spike glycoprotein (S protein) is a transmembrane protein with a molecular weight of about 150 kDa, forms homotrimers stand out in the viral surface. S protein facilitates the binding of envelope viruses to Angiotensin-Converting Enzyme 2 (ACE-2) receptors in lower respiratory tract cells. S protein consist of 2 part, S1 and S2. S1 is responsible for the determination of the host-virus range and cellular tropism with the receptor-binding domain make-up while S2 functions to mediate virus fusion in transmitting host cells^{18,16}

SARS-CoV-2 rapidly is transmitted from human to human pathogen and causes a wide spectrum of clinical manifestations in COVID-19 patients. The major clinical manifestation of

COVID-19 is similar to other human CoV infection. In general, SARS-CoV-2 is associated with high viral loads in upper respiratory tract secretion. COVID-19 is developed into Severe Acute Respiratory Infections and Acute Distress Syndrome especially in elderly, comorbidities, and immunosuppression patient.¹⁹

Human to human transmission of COVID-19 mainly occurs via respiratory droplets when patients with coughs, sneezes, or talks can infect another person via direct contact with the mucous membranes. The infection can also occur if a person touches an infected surface and then touches their eye, nose, or mouth. The droplet does not travel that 2 meters and not linger than 3 hours in the air.²⁰

Based on the presence of clinical features, COVID-19 is divided into 2 groups such as asymptomatic and symptomatic infection. Asymptomatic COVID-19 infection shows positive SARS-CoV-2 nucleic acid testing of the pharyngeal swab samples but does not show the clinical feature until the first day of continuous negative tests (1-21 days). Only 20.8% asymptomatic cases develop into symptoms cases during hospitalization such as fever, cough, fatigue and etc, 50% cases showed typical CT images of the ground-glass chest, and five (20.8%) presented stripe shadowing in the lungs. The remaining 29.2% cases showed a normal CT image and had no symptoms during hospitalization (young patient estimated 14 years). However, no one cases develop into severe COVID-19 pneumonia or died. Asymptomatic carrier patient has the potential to transmit the disease to others. This study also shows that Asymptomatic COVID-19 pneumonia.²¹

According to some studies, an incubation period of COVID-19 is 5 – 14 days following exposure and cause the respiratory and systemic disorder.¹³ The spectrum of symptomatic COVID-19 infection ranges from mild to critical. The mild stage is signed with high fever, dry cough, headache, fatigue, sputum production, lymphopenia, and diarrhea. COVID-19 infection cause cough (days 1 – 14), chills (days 1 – 9), high fever (days 7 – 9), fatigue (days 9 – 14), and shortness of breath (days 9-14). The severe stage is signed with dyspnea and hypoxia. The pathological finding of COVID-19 associated with acute respiratory disruption syndrome. The critical disease is signed with respiratory failure, shock, or multi-organ dysfunction. COVID-19 infection causes acute respiratory distress syndrome (ARDS) until death.⁶

The infection show laboratory features such as lymphopenia and elevated some molecules (liver enzymes, lactate dehydrogenase, elevated inflammatory markers, D-dimer, prothrombin time, troponin, creatine phosphokinase, and acute kidney injury. The laboratory testing shows positive in Viral RNA PCR testing and Immunoglobulin M (IgM), and there is ground-glass chest, stripe shadowing in the lungs in CT Scan, and pneumonia in x-ray test.^{22,23}

COVID-19 is a positive-sense single-stranded RNA (ssRNA) that entry to the patient via respiratory droplets when patients with coughs, sneezes, or talks can infect another person via direct contact with the mucous membranes and touching the infected surface the body (eye, nose or mouth).

COVID-19 S protein (envelope spike glycoprotein) will bind to ACE2 for SARS-CoV,¹⁹ ACE2 SARS-CoV-2,²⁴ CD209L (a C lectin or called L-SIGN) for SARS-CoV,²⁵ DPP4 for MERS-CoV²⁶ then occur fusion between the viral cell membrane and the target cell.²⁷ Proteolytic cleavage occurs in S SARS-CoV protein in position (S2') mediated by membrane fusion and virus infectivity.²⁸ MERS-CoV has also evolved through the activation of two abnormal steps for furin membrane fusion.²⁹ In addition to membrane fusion, clathrin-dependent and clathrin-independent endocytosis also mediate the process of SARS-CoV virus entry into cells.^{30,31}

After that viral RNA enters the cytoplasm of cells in the form of polyproteins and structural proteins which then replicates into new viral RNA. The cell will then be controlled by the structural protein SARS-CoV to form an envelope protein that will be inserted into the endoplasmic reticulum (ER) and Golgi apparatus while the nucleocapsid is formed by a combination of viral RNA genomes and nucleocapsid proteins. In the end, the virus particles will be brought to endoplasmic reticulum-Golgi intermediate compartment (ERGIC) for maturation and will then be removed by ERGIC with vesicles containing infectious viruses

through vesicle vesicles which will docking with membranes so that the virus will be released again and ready to infect other healthy cells.¹⁹

Viral antigen entry stimulates an immune response. COVID-19 viral antigen will be presented to the antigen-presenting cell (APC). The antigenic peptide will be presented by MHC (human leukocyte antigen or HLA in humans) to the specific cytotoxic T lymphocyte virus. MHC Class I played the most role in this Ag COVID-19 presentation, although MHC class II also played a role.⁸ The COVID-19 presentation antigen was not fully understood but can be associated with antigen presentation on SARS-CoV or MERS-CoV. Certain HLA polymorphisms have a risk of being susceptible to SARS-CoV infection, namely HLA-B * 4601, HLA-B * 0703, HLA-DR B1 * 1202⁹ and HLA-Cw * 0801,¹⁰ while HLA-DR0301, HLA-Cw 1502 and HLA-A * 0201 has protection against SARS infection.¹¹ MHC class II molecules cause susceptibility to MERS-CoV infection,¹³ such as HLA-DRB1 * 11: 01 and HLA-DQB1 * 02: 0. In addition, the Mannose Binding Lectine (MBL) gene polymorphism associated with antigen presentation is associated with susceptibility to SARS-CoV infection.¹⁴

SARS-CoV-2 stimulate innate and adaptive immune responses. SARS-CoV-2 leads to infection of ACE2 expressing target cells such as alveolar type 2 cells or other cell in lung. SARS-CoV-2 causes an increase in total neutrophils, reduces total lymphocytes, increases serum IL-6, and increases C-reactive protein. Increased neutrophils and decreased lymphocytes also correlate with disease severity and death. COVID-19 has higher plasma levels of many pro inflammatory cytokines, IP-10, MCP-1, MIP-1A, and TNFα.2 that show highly pro-inflammatory conditions in the disease progression and severity. Activation of neutrophil as Antigen Presenting Cell (APC) increase Th1/Th17 expression. Th1/Th17 is induced adaptive response by increase specific antibodies production. Viral antigen may decrease antiviral IFN responses resulting in uncontrolled viral replication whereas IFN is effective innate immune response against viral infection and induce effective adaptive immune response. Type I IFN suppression and hyper pro-inflammatory cytokine production be the basis of viral infection treatment including SARS-CoV infection.¹⁵

In adaptive immune response, COVID-19 viral antigen will stimulate humoral and cell mediated immunity by B cells and T cells. B cells will form IgM and IgG anti SARS-CoV. IgM will disappear on the 12th day while IgG will last a long time in the body which will function as a protector against infection with the same virus strain.¹⁶ IgG anti SARS-CoV is S-specific and N-specific.¹⁹ In general, Th1 plays a dominant role in an adaptive immunity against to viral infections. Specific Th1/Th17 may be activated and contributes to exacerbate inflammatory responses. B cells/plasma cells produce SARS-CoV-2 specific antibodies that may help neutralize viruses. In previous study showed the peak of specific IgM at day 9 after disease onset and the switching to IgG by week 2. Whether the titer of specific antibody correlates with disease severity remains to be investigated.¹⁵

In cellular immune response, the number of CD4⁺ and CD8⁺ cellular immunity in peripheral blood will drop significantly in SARS-CoV-2 infection.¹⁷ Patients recovering from SARS-CoV will have CD4⁺ and CD8⁺ memory T cells that last for 4 years so that when there is repeated viral exposure the body will be able to produce IFN-4.³² In some patients the specific T cell memory of SARS-CoV can still be identified.¹² CD8 + T cells specific to MERS CoV are also found in mice that are infected with the virus.¹² In general, CD8+ T cell responses are more frequent with greater magnitude than CD4+ T cell responses in SARS Coronavirus. Strong T cell responses correlated significantly with higher neutralizing antibody while more serum Th2 cytokines (IL-4, IL-5, IL-10). In MERS-CoV infection, early rise of CD8+ T cells correlates with disease severity and at the convalescent phase, dominant Th1 type helper T cells are observed. In an animal model, airway memory CD4+ T cells specific for conserved epitope are protective against lethal challenge and can cross react with SARS-CoV and MERS-CoV. Th1 type and CD8+ T cell are keys for successful control of SARS-CoV and MERS-CoV and probably true for SARS-CoV-2 as well to suppress lung pathogenesis.¹⁵

The leading cause of death in COVID-19 infection is Acute Respiratory Distress Syndrome (ARDS) and multi-organ dysfunction. ARDS is the leading cause of death. This ARDS is an immuno-pathological event from SARS-Cov-2, SARS-CoV and MERS-CoV infections.¹⁶ More patients also developed lymphopenia and pneumonia with characteristic pulmonary ground

glass opacity changes on chest CT. COVID-19 infection also show cytokine storm. Cytokine storm is inflammation that cannot be controlled and is very deadly due to the body's response due to the large production of inflammatory cytokines such as (IFN- α , IFN- γ , IL-1 β , IL-6, IL-12, IL-18, IL-33, TNF- α , TGF- β etc.), as well as some chemokines such as (CCL2, CCL3, CCL5, CXCL8, CXCL9, CXCL10 etc) produced by several immune effector cells produced by SARS-CoV infection.^{33,34,35} For example in patients with severe MERS-CoV infection, an increase in the production of cytokines IL-6, IFN- α , CCL5, CCL8 and CXCL-10 in their serum compared with patients who have mild to moderate symptoms.³⁶ The production of pro-inflammatory cytokines mentioned above causes the patient's body to experience ARDS which can cause multiple organ failure (MOF) and ARDS until death. The presence of lymphopenia and cytokine storm may important role in COVID-19 pathogenesis. The Cytokine storm can initiate viral sepsis and inflammatory-induced lung injury which lead to other complications including pneumonitis, ARDS, respiratory failure, shock, multi-organ failure and potentially death.¹⁷

SARS-CoV and MERS-CoV use various strategies in order to survive against host immunity. Microbes have pathogen-associated molecular patterns (PAMPs) that can be recognized by pattern recognition receptors (PRRs). SARS-CoV and MERS-CoV can induce the production of double membrane vesicles containing little PRRs and can replicate in these vesicles, so that SARS-CoV dsRNA can avoid detection of the host immune system.³⁷ Hosts can produce IFN- α and IFN- β which can protect the effects of SARS-CoV infection.^{38,39} Another corona virus, MERS-CoV, has a protein accessory molecule 4a that can inhibit MDA5 activation to induce IFN production by interacting directly with dsRNA.⁴⁰ MERS-CoV also has ORF4a, ORF4b, ORF5 and viral protein membranes that can inhibit the transport of IFN regulatory factor 3 (IRF3) and inhibit the activation of IFN- β promoters.⁴¹ MERS CoV can also hold genes that play a role in the presentation of Ag viruses.⁴²

4. CONCLUSION

Specific studies on the immune response to SARS-CoV-2 are still developing. Various study approaches can be carried out by looking at the clinical manifestations that emerge, the results of examinations and literature review from their predecessors, namely SARS-CoV and MERS-CoV. So far, it can be concluded that molecular studies related to SARS-CoV-2 immunopathology states that SARS-CoV-2 increases pro-inflammatory cytokine production, suppresses IFN, increases neutrophil count, increases Th1/Th17 expression, B cell response, and CD8+ and CD4+ responses. Cytokine storms are known to be a trigger for serious respiratory problems and multi-organ damage that can cause death.

DISCLOSURE STATEMENT

No potential conflict of interest was reported by the authors.

ACKNOWLEDGEMENT

We would like thank to STIKES Rumah Sakit Anwar Medika, Jurnal Teknologi Laboratorium (Journal of Laboratory Technology), editor and reviewer for editing the manuscript.

FUNDING INFORMATION

STIKES Rumah Sakit Anwar Medika, Sidoarjo, Indonesia.

REFERENCES

- 1. Lu H, Stratton CW, Tang YW. Outbreak of pneumonia of unknown etiology in Wuhan, China: The mystery and the miracle. *J Med Virol*. 2020;92(4):401-402. doi:10.1002/jmv.25678
- 2. World Health Organization. *Novel Coronavirus (2019-NCoV) Situation Report 12.* World; 2020. <u>https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200201-sitrep-12-ncov.pdf?sfvrsn=273c5d35_2</u>.
- 3. Bassetti M, Vena A, Giacobbe DR. The novel Chinese coronavirus (2019-nCoV) infections: Challenges for fighting the storm. *Eur J Clin Invest*. 2020;50(13209):1-4.

doi:<u>10.1111/eci.13209</u>

- 4. World Health Organization. *Coronavirus Disease 2019 (COVID-19) Situation Report 70.* World; 2020. <u>https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200330-sitrep-70-covid-19.pdf?sfvrsn=7e0fe3f8_4</u>.
- 5. Kementerian Kesehatan Repulik Indonesia. *Situasi COVID-19*. Indonesia; 2020. <u>https://www.kemkes.go.id/</u>.
- Li Q, Guan X, Wu P, et al. Early transmission dynamics in Wuhan, China, of novel coronavirus-infected pneumonia. N Engl J Med. 2020;382(13):1199-1207. doi:10.1056/NEJMoa2001316
- 7. Wang W, Tang J, Wei F. Updated understanding of the outbreak of 2019 novel coronavirus (2019-nCoV) in Wuhan, China. *J Med Virol*. 2020;92(4):441-447. doi:10.1002/jmv.25689
- Liu J, Wu P, Gao F, et al. Novel immunodominant peptide presentation strategy: a featured HLA-A*2402-restricted cytotoxic T-lymphocyte epitope stabilized by intrachain hydrogen bonds from severe acute respiratory syndrome coronavirus nucleocapsid protein. *J Virol.* 2010;84(22):11849-11857. doi:10.1128/jvi.01464-10
- 9. Keicho N, Itoyama S, Kashiwase K, et al. Association of human leukocyte antigen class II alleles with severe acute respiratory syndrome in the Vietnamese population. *Hum Immunol.* 2009;70(7):527-531. doi:10.1016/j.humimm.2009.05.006
- 10. Chen YMA, Liang SY, Shih YP, et al. Epidemiological and genetic correlates of severe acute respiratory syndrome coronavirus infection in the hospital with the highest nosocomial infection rate in Taiwan in 2003. *J Clin Microbiol*. 2006;44(2):359-365. doi:10.1128/JCM.44.2.359-365.2006
- 11. Wang S-F, Chen K-H, Chen M, et al. Human-leukocyte antigen class I CW 1502 and class II DR 0301 genotypes are associated with resistance to severe acute respiratory syndrome (SARS) infection. *Viral Immunol.* 2011;24(5). doi:<u>10.1089/vim.2011.0024</u>
- 12. Tang F, Quan Y, Xin Z-T, et al. Lack of peripheral memory B cell responses in recovered patients with severe acute respiratory syndrome: a six-year follow-up study. *J Immunol.* 2011;186(12):7264-7268. doi:10.4049/jimmunol.0903490
- 13. Hajeer AH, Balkhy H, Johani S, Yousef MZ, Arabi Y. Association of human leukocyte antigen class II alleles with severe Middle East respiratory syndrome-coronavirus infection. *Ann Thorac Med.* 2016;11(3):211-213. doi:10.4103/1817-1737.185756
- 14. Tu X, Chong WP, Zhai Y, et al. Functional polymorphisms of the CCL2 and MBL genes cumulatively increase susceptibility to severe acute respiratory syndrome coronavirus infection. *J Infect*. 2020;71(1):101-109. doi:10.1016/j.jinf.2015.03.006
- 15. Prompetchara E, Ketloy C, Palaga T. Immune responses in COVID-19 and potential vaccines: lessons learned from SARS and MERS epidemic. *Asian Pacific J Allergy Immunol.* 2020. doi:10.12932/AP-200220-0772
- 16. Li G, Fan Y, Lai Y, et al. Coronavirus infections and immune responses. *J Med Virol*. 2020;92(4):424-432. doi:10.1002/jmv.25685
- 17. Xu Z, Shi L, Wang Y, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respir Med.* 2020;8(4):420-422. doi:10.1016/S2213-2600(20)30076-X
- 18. Su S, Wong G, Liu J, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. *Trend Microbiol*. 2016;24(6):490-502. doi:10.1016/j.tim.2016.03.003
- 19. Guan W, Ni Z, Hu Y, et al. Clinical characteristics of 2019 novel coronavirus infection in China. *N Engl J Med*. 2020;382:1708-1720. doi:10.1056/NEJMoa2002032
- 20. Doremalen N van, Bushmaker T, Morris DH, et al. Aerosol and surface stability of SARS-CoV-2 as compared with SARS-CoV-1. *N Engl J Med.* 2020;382:1564-1567. doi:10.1056/NEJMc2004973
- Hu Z, Song C, Xu C, et al. Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China. Sci China Life Sci. 2020;63(5):706-711. doi:10.1007/s11427-020-1661-4
- 22. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet.* 2020;395(10223):497-506. doi:10.1016/S0140-

64

PERSPECTIVE OF IMMUNE RESPONSE

6736(20)30183-5

- 23. Zhou F, Yu T, Du R, et al. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet.* 2020;395(10229):1054-1062. doi:10.1016/S0140-6736(20)30566-3
- 24. De Wit E, Van Doremalen N, Falzarano D, Munster VJ. SARS and MERS: recent insights into emerging coronaviruses. *Nat Rev Microbiol*. 2016;14(8):523-534. doi:10.1038/nrmicro.2016.81
- 25. Jeffers SA, Tusell SM, Gillim-Ross L, et al. CD209L (L-SIGN) is a receptor for severe acute respiratory syndrome coronavirus. *Proc Natl Acad Sci U S A*. 2004;101(44):15748-15753. doi:<u>10.1073/pnas.0403812101</u>
- 26. Raj VS, Mou H, Smits SL, et al. Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature*. 2013;495(7440):251-254. doi:10.1038/nature12005
- 27. Simmons G, Reeves JD, Rennekamp AJ, Amberg SM, Piefer AJ, Bates P. Characterization of severe acute respiratory syndrome-associated coronavirus (SARS-CoV) spike glycoprotein-mediated viral entry. *Proc Natl Acad Sci U S A*. 2004;101(12):4240-4245. doi:10.1073/pnas.0306446101
- 28. Belouzard S, Chu VC, Whittaker GR. Activation of the SARS coronavirus spike protein via sequential proteolytic cleavage at two distinct sites. *Proc Natl Acad Sci U S A*. 2009;106(14):5871-5876. doi:10.1073/pnas.0809524106
- 29. Mille JK, Whittaker GR. Host cell entry of Middle East respiratory syndrome coronavirus after two-step, furin-mediated activation of the spike protein. *Proc Natl Acad Sci U S A*. 2014;111(42):15214-15219. doi:<u>10.1073/pnas.1407087111</u>
- 30. Wang H, Yang P, Liu K, et al. SARS coronavirus entry into host cells through a novel clathrin- and caveolae-independent endocytic pathway. *Cell Res.* 2008;18(2):290-301. doi:10.1038/cr.2008.15
- 31. Kuba K, Imai Y, Ohto-Nakanishi T, Penninger JM. Trilogy of ACE2: A peptidase in the renin-angiotensin system, a SARS receptor, and a partner for amino acid transporters. *Pharmacol Ther.* 2010;128(1):119-128. doi:<u>10.1016/j.pharmthera.2010.06.003</u>
- 32. Fan YY, Huang ZT, Li L, et al. Characterization of SARS-CoV-specific memory T cells from recovered individuals 4 years after infection. *Arch Virol.* 2009;154(7):1093-1099. doi:10.1007/s00705-009-0409-6
- 33. Williams AE, Chambers RC. The mercurial nature of neutrophils: Still an enigma in ARDS? *Am J Physiol Lung Cell Mol Physiol*. 2014;306(3):L217-L230. doi:10.1152/ajplung.00311.2013
- 34. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. *Semin Immunopathol.* 2017;39(5):529-539. doi:10.1007/s00281-017-0629-x
- 35. Cameron MJ, Bermejo-Martin JF, Danesh A, Muller MP, Kelvin DJ. Human immunopathogenesis of severe acute respiratory syndrome (SARS). *Virus Res.* 2008;133(1):13-19. doi:10.1016/j.virusres.2007.02.014
- 36. Min CK, Cheon S, Ha NY, et al. Comparative and kinetic analysis of viral shedding and immunological responses in MERS patients representing a broad spectrum of disease severity. *Sci Rep.* 2016;6:1-12. doi:<u>10.1038/srep25359</u>
- 37. Snijder EJ, van der Meer Y, Zevenhoven-Dobbe J, et al. Ultrastructure and Origin of Membrane Vesicles Associated with the Severe Acute Respiratory Syndrome Coronavirus Replication Complex. *J Virol.* 2006;80(12):5927-5940. doi:10.1128/jvi.02501-05
- 38. Channappanavar R, Fehr AR, Vijay R, et al. Dysregulated type I interferon and inflammatory monocyte-macrophage responses cause lethal pneumonia in SARS-CoV-infected mice. *Cell Host Microbe*. 2016;19(2):181-193. doi:10.1016/j.chom.2016.01.007
- 39. Channappanavar R, Fehr AR, Zheng J, et al. IFN-I response timing relative to virus replication determines MERS coronavirus infection outcomes. *J Clin Invest.* 2019;129(9):3625-3639. doi:<u>10.1172/JCI126363</u>

65

- 40. Niemeyer D, Zillinger T, Muth D, et al. Middle east respiratory syndrome coronavirus accessory protein 4a is a type I interferon antagonist. *J Virol.* 2013;87(22):12489-12495. doi:10.1128/JVI.01845-13
- 41. Yang Y, Zhang L, Geng H, et al. The structural and accessory proteins M, ORF 4a, ORF 4b, and ORF 5 of Middle East respiratory syndrome coronavirus (MERS-CoV) are potent interferon antagonists. *Protein and Cells*. 2013;4:951–961. doi:<u>10.1007/s13238-013-3096-8</u>
- 42. Menachery VD, Schäfer A, Burnum-Johnson KE, et al. MERS-CoV and H5N1 influenza virus antagonize antigen presentation by altering the epigenetic landscape. *Proc Natl Acad Sci U S A*. 2018;115(5):E1012-E1021. doi:<u>10.1073/pnas.1706928115</u>

SHORT BIOGRAPHY



Martina Kurnia Rohmah has completed her Master's Degree in 2015 at the Medical Faculty, Brawijaya University and has completed Master Student Exchange Program at Graduated School of Science and Technology Kumamoto Univesity, Japan. She worked as a lecturer at Departement of Pharmacy, STIKES Rumah Sakit Anwar Medika. Her research area focused on Biomedical and Molecular Science.



Arif Rahman Nurdianto has completed his Doctoral Degree in 2020 at the Medical Faculty, Airlangga University. He worked as a doctor at Public Health Office of Trosobo, Sidoarjo and as a lecturer at Departement of Medical Laboratory Technology, STIKES Rumah Sakit Anwar Medika. His research area focused on Immunology and Parasitology.